

Computational structure-activity relationship (SAR) of berberine analogs in double-stranded and G-quadruplex DNA binding reveals both position and target dependence

Stephanie Sun¹, Bhavesh Ashok², Andrew Su³, Saira Hamid⁴, Karthikha Sri Indran⁴, Aashi Shah², Sarah Su⁵, Simrun Sakhrani⁶, Edward Njoo⁷

¹BASIS Independent Silicon Valley, San Jose, California

²Amador Valley High School, Pleasanton, California

³Foothill High School, Pleasanton, California

⁴Mission San Jose High School, Fremont, California

⁵Los Altos High School, Los Altos, California

⁶The College Preparatory School, Oakland, California

⁷Department of Chemistry, Biochemistry, & Physical Science, Fremont, California

SUMMARY

Berberine, a natural product alkaloid, and its analogs have a wide range of medicinal properties, including antibacterial and anticancer effects. Previous studies showed that berberine and its analogs intercalate into DNA, thereby inhibiting DNA replication. Berberine has also been studied as a photosensitizer and was shown to generate reactive singlet oxygen *in situ*, and this has applications in photodynamic therapy. Various groups have synthesized berberine analogs that have comparable or improved biological activity; however, an exhaustive structure-activity relationship of the free energy of binding of berberine analogs with substitution at C-8, C-12, and C-13 to DNA have not been previously reported. High-throughput virtual screening (HTVS) allows for efficient analysis of compound libraries to identify lead compounds as possible pharmaceutical agents. Here, we employed HTVS towards a library of alkyl or aryl berberine analogs on C-8, C-12, and C-13 to probe binding to double-stranded and G-quadruplex DNA. Predicted free energies of binding to double-stranded DNA and G-quadruplex DNA were generated via molecular docking. The excited state electronic structure calculations were conducted via time-dependent density functional theory to probe the potential photosensitizing activity of each compound, and the potential G-quadruplex stabilizing abilities of key berberine analogs were probed through molecular dynamics simulations on a 4.0 nanosecond timescale. We determined that the nature of the substituent, the position of the substituent, and the nucleic acid target affect the free energy of binding of berberine analogs to DNA and G-quadruplex DNA, however berberine analogs did not result in net stabilization of G-quadruplex DNA.

INTRODUCTION

Berberine (Figure 1), a naturally occurring isoquinoline alkaloid extracted from the roots and stem of plants from the genus *Berberis*, has been of great medicinal interest due to its wide range of reported biological activities, including antimicrobial, antidiabetic, and anticancer activity (1-5). Berberine-containing extracts have been used as a medicinal agent in many traditional cultures dating back to 3000 BC (6).

The mechanism of action of berberine is believed to be intercalation with DNA, where it acts as a photosensitizer. Berberine was previously found to intercalate in DNA with a high free energy of binding (ΔG), driven by electrostatic attractions and pi-stack interactions between the compound and nucleotide base pairs in DNA secondary structures (7). Upon photoirradiation of the berberine-DNA complex, berberine acts as a photosensitizer and generates singlet oxygen, a reactive oxygen species, which oxidizes guanines and results in DNA damage, thereby inhibiting DNA replication and halting cell division (8). This was previously studied in application to photodynamic therapy as a potential treatment of various types of cancers and antimicrobial agents (9-10). Berberine has also been reported to stabilize G-quadruplex DNA (G4DNA), a four-stranded, noncanonical secondary structure of DNA consisting of guanine-rich DNA sequences

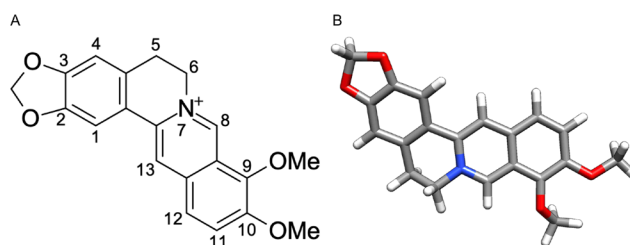


Figure 1. Background and introduction of Berberine 1.

(a) Chemical structure of berberine with the carbons numbered.

(b) 3D structure of berberine (DFT optimized, B3LYP, def2-SVP).

(11). Stabilization of G4DNA inhibits telomerase, an enzyme that is overexpressed in cancers; this inhibition of telomerase results in the inhibition of cancer activity (12).

Several semisynthetic analogs and derivatives of berberine have previously been prepared and evaluated for biological activities; some of these are reported to have comparable or superior antibacterial, antifungal, or anticancer activity compared to the natural product, and some have been reported to possess improved free energies of binding to DNA and G4DNA. Addition of an alkyl or aryl chain to C-8 of berberine can be achieved via nucleophilic addition of alkyl or aryl Grignard; such compounds have been reported to have more potent antimicrobial activity (13), but the possible role of the stereogenic center at C-8 in DNA binding is not yet known (Figure 2a). Wang *et al.* previously reported a library of 12-amine-berberine derivatives that demonstrate increased anticancer activity compared to berberine (14). The synthesis and biological screening of 13-alkylberberine analogs revealed that the addition of alkyl chains of various lengths

improves the anticancer, anti-inflammatory, and antioxidant activity of berberine (15-16). It has also been reported that a borohydride reduction at the C-8 iminium to yield dihydroberberine generally results in a loss of antimicrobial efficacy (17). While many have studied the biological activities of berberine and its analogs, less than 20 percent of studies reported in the last 10 years quantify the DNA free energy of binding of such analogs, and even fewer have produced crystal structures of berberine bound to a DNA target.

Here, we report an extensive *in silico* virtual screen of a representative library of 8-, 12-, and 13-alkyl and aryl berberine analogs (Figure 2b). The use of HTVS, which screens small molecule libraries against potential drug targets, has enabled the rapid and efficient screening of large libraries of chemical entities (18-20). We employed molecular docking to determine the predicted free energy of binding of each analog to each of the two DNA targets and we performed molecular dynamics (MD) simulations on berberine and selected analogs to simulate potential ligand-mediated stabilization

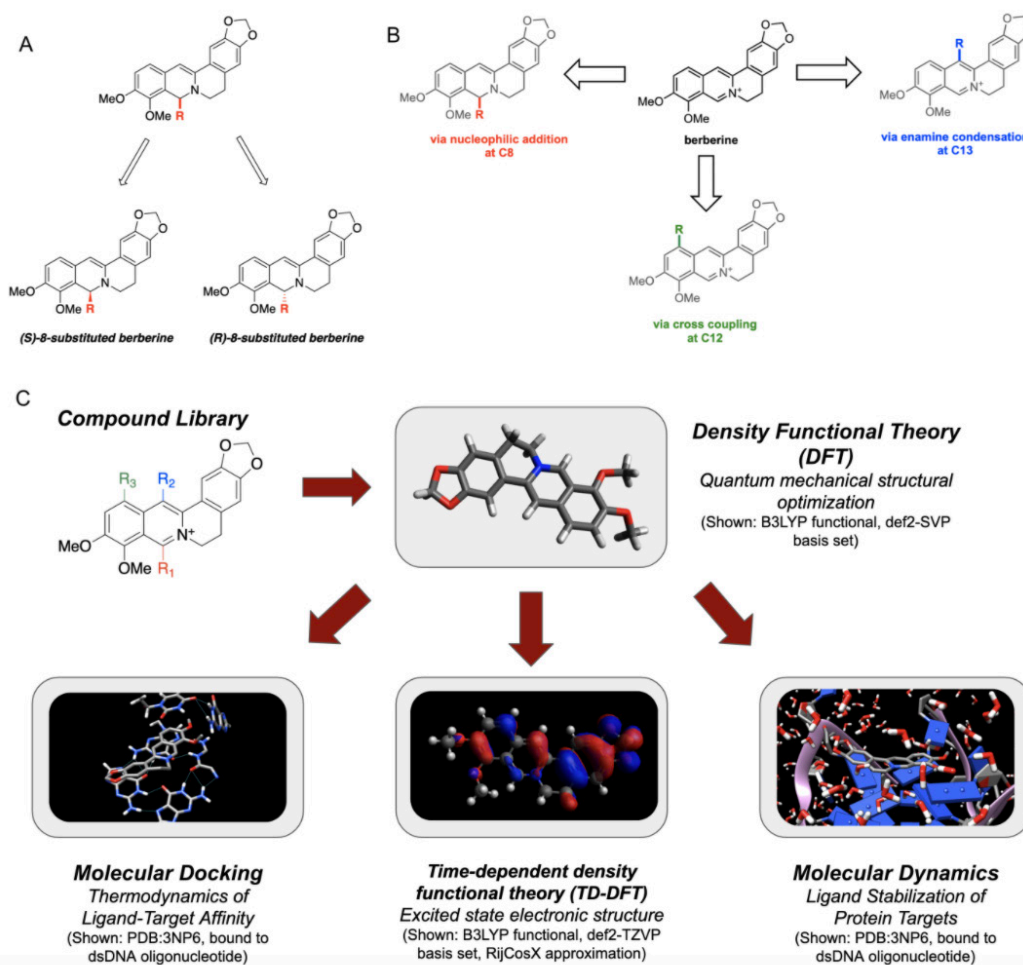
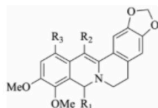


Figure 2. Design of the library of berberine analogs and our methodology in this study. (a) A stereocenter is formed when alkyl and aryl chains are added to C-8 of berberine, resulting in R and S enantiomers. Both R and S enantiomers of all C-8 analogs were studied. **(b)** Possible reactions for the synthesis of C-8, C-12, and C-13 analogs that inspired the design of the library. Possible reactions include a nucleophilic addition to C-8 with Grignard reagents, treatment of berberine with elemental bromine yields 12-bromoberberine, which can serve as a handle for cross coupling reactions, and enamine condensations to C-13. **(c)** Workflow and methodology in our work.

of the G4DNA complex. We implemented time-dependent density functional theory (TD-DFT) calculations to predict the wavelength of maximal absorbance and the relative energies of the singlet and triplet excited states of each analog. The energetic ordering of these excited states affects whether the berberine analogs can generate singlet oxygen, the process of which can result in DNA damage (Figure 2c). Since many DNA intercalators rely on pi-stack interactions or electrostatic attractions, we initially hypothesized that the addition of aromatic systems to C-8, C-12, or C-13 would provide an increase in ΔG , and that loss of the cationic iminium at C-8 would diminish the free energy of binding (21).

In this work, we screened a library of 31 berberine analogs against 2 biological targets, double-stranded DNA (dsDNA) d(CGTCAG) and G4DNA, based on previously-reported crystal structures (PDB codes 3NP6 for dsDNA and 6JWD for G4DNA) (22-23). The results of the HTVS, MD simulations, and TD-DFT calculations indicate that the impact of aryl and aliphatic substitution on DNA free energy of binding is both position-dependent and target-dependent. We determined that the binding affinity of berberine analogs to dsDNA and G4DNA is affected by the nature and carbon position of the substituent, and these compounds have no net stabilizing effect on G4DNA complexes.



Compound 2, 8a-h

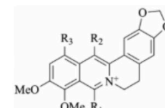
Compound	R ₁	R ₂	R ₃	ΔG - dsDNA	ΔG - G4DNA
2	H	H	H	-6.2	2.5
8a	CH ₃	H	H	R = -4.0, S = -4.1	R = -0.1, S = 2.3
8b	C ₂ H ₅	H	H	R = -4.2, S = -3.8	R = 0.8, S = 1.4
8c	C ₄ H ₉	H	H	R = -4.5, S = -4.3	R = 3.3, S = 5.0
8d	C ₆ H ₁₃	H	H	R = -3.8, S = -3.6	R = 3.0, S = 2.5
8e	C ₈ H ₁₇	H	H	R = -3.7, S = -3.6	R = 6.3, S = 1.5
8f	C ₁₀ H ₂₁	H	H	R = -3.0, S = -3.2	R = 9.1, S = 3.4
8g		H	H	R = -4.4, S = -3.4	R = 6.1, S = 9.8
8h		H	H	R = -2.1, S = -3.3	R = 9.3, S = 9.2
8i		H	H	R = -1.0, S = -2.2	R = 65.9, S = N/A

RESULTS

Molecular docking of the library of berberine analogs with dsDNA d(CGTCAG) and G4DNA was completed using Autodock Vina (24-25). Docking parameters were determined from previously reported crystal structures, and the high degree of similarity in ligand positioning achieved between docked poses and the crystal structure demonstrates the predictivity of the docking parameters used (22-23). Results were quantified based on the ΔG in kcal/mol (Figure 3).

dsDNA Binding

Docking results for analogs with bromo-, phenyl-, and naphthyl- substitutions at the C-12 position had greater ΔG to dsDNA than analogs with substitutions at C-8 and C-13, and some of these compounds exhibited greater ΔG values than berberine itself. A lower ΔG value indicates stronger binding affinity to target proteins, and a negative ΔG indicates that the interaction between the compound and target is thermodynamically stable. Compounds with naphthyl groups on C-12 had a ΔG of -6.0 and -6.8 kcal/mol (Figure 4c), compound 12c and 12d, respectively. Interestingly, naphthyl additions to the C-8 position yielded lower ΔG values than berberine, with ΔG values of -2.1 and -3.3 kcal/mol for the R and S enantiomers of compound 8h, respectively. The same trend was seen in the R and S enantiomers of compound 8i, whose ΔG increased to -1.0 kcal/mol and -2.2 kcal/mol, respectively.



Compound 1, 12a-d, 13a-g

Compound	R ₁	R ₂	R ₃	ΔG - dsDNA	ΔG - G4DNA
1	H	H	H	-5.6	2.6
12a	H	H	Br	-5.8	4.0
12b	H	H		-5.9	25.7
12c	H	H		-6.0	35.5
12d	H	H		-6.8	N/A
13a	H	C ₂ H ₅	H	-4.8	3.2
13b	H	C ₆ H ₁₁	H	-4.8	7.4
13c	H	C ₈ H ₁₇	H	-4.6	4.1
13d	H	C ₁₀ H ₂₁	H	-4.3	8.2
13e	H		H	-4.4	10.4
13f	H		H	-4.9	4.9
13g	H		H	-4.2	11.1

Figure 3. Thermodynamics of the binding of the library berberine analogs to dsDNA and G4DNA. Free energies of binding (ΔG) are reported in kcal/mol and represent the ΔG of the most thermodynamically-stable predicted binding pose.

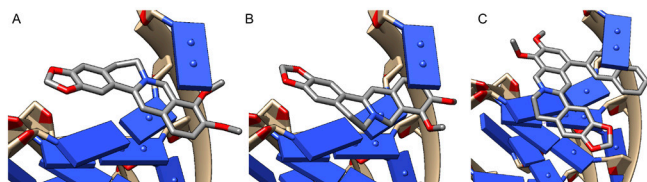


Figure 4. Most thermodynamically stable binding pose of representative compounds in our library to dsDNA. (a) Berberine 1 ($\Delta G = -5.6$ kcal/mol) (b) Dihydroberberine 2 ($\Delta G = -6.2$ kcal/mol) (c) Compound 12d, the berberine analog with the best free energy of binding to dsDNA ($\Delta G = -6.8$ kcal/mol).

G4DNA Binding

Berberine analogs had lower free energies of binding to G4DNA than dsDNA. Unlike dsDNA, aliphatic chains on the C-8 positions demonstrated the best binding affinity to G4DNA. The R enantiomer of compound **8a** had the lowest ΔG value of all analogs screened at -0.1 kcal/mol (**Figure 5c**). Additions of naphthyl groups at both C-8 and C-12 greatly decreased the ΔG values of analogs **8h**, **8i**, **12c**, and **12d** to G4DNA. Compounds **8i** and **12d** were not able to dock to G4DNA. It is unclear why AutoDock Vina returned positive ΔG values for the DNA-berberine binding interaction.

The lengthening of the alkyl chain on the C-8 analogs also resulted in decreased free energy of binding of the compound to G4DNA. This could be attributed to steric clashes, in which members of the alkyl chain past the fourth carbon overlap with atoms of the receptor without taking part in the binding interactions. This is thermodynamically unfavorable due to repulsive electron interactions between these clashing atoms, increasing the ΔG value. Differences were also observed between the R and S enantiomers of these analogs, with thermodynamic favorability of the R enantiomer for alkyl chains four carbons and shorter. This trend seems to be

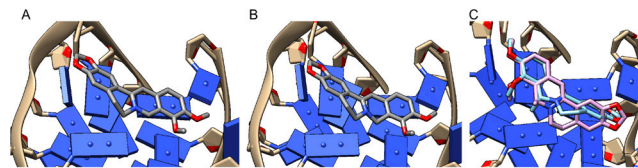


Figure 5. Binding poses of berberine and berberine analogs to G4DNA. (a) Berberine 1 ($\Delta G = 2.6$ kcal/mol) (b) Dihydroberberine 2 ($\Delta G = 2.5$ kcal/mol) (c) Both enantiomers of compound **8a**. The structure in blue is the R enantiomer ($\Delta G = -0.1$ kcal/mol), and the structure in purple is the S enantiomer ($\Delta G = 2.3$ kcal/mol).

reversed for alkyl chains longer than four carbons, with the S enantiomer having lower ΔG values.

Time-Dependent Density Functional Theory

TD-DFT was utilized to study the excited state electronic structure of each compound. Moreover, the molecular orbital energies that resulted allowed us to probe whether berberine analogs could undergo electronic transitions that can effectively produce singlet oxygen (**Figure 6**).

Here, we found that the energy gap generated by berberine and all analogs screened was sufficient for the production of singlet oxygen species. The energy emitted through phosphorescence, the transition from the triplet excited state back to ground state, is transferred to oxygen for the production of singlet oxygen (**Figure 7**). The energy gap necessary for the production of the singlet oxygen is 0.98 eV, which all of our compounds are able to undergo (26). The absorbance maximum wavelengths of the analogs of berberine undergo a blueshift. The C-8 compounds, which contain the reduced isoquinoline chromophore, undergo a larger blueshift than analogs with the same isoquinolinium chromophore as berberine. The rate of intersystem crossing

Compound	E_{S_0} (eV)	E_{S_1} (eV)	E_{T_1} (eV)	$\Delta E_{S_0 \rightarrow S_1}$ (H \rightarrow L)	$\Delta E_{T_1 \rightarrow S_0}$ (H \rightarrow L)	λ_{max} (nm)
1	-5.645	-2.701	-3.854	2.944	1.791	421
2	-4.735	-1.119	-2.447	3.616	2.288	343
8a	-4.775	-1.069	-2.400	3.706	2.375	335
8b	-4.768	-1.048	-2.382	3.720	2.396	334
8c	-4.688	-1.123	-2.447	3.565	2.241	348
8d	-4.685	-1.112	-2.430	3.573	2.255	347
8e	-4.861	-1.117	-2.436	3.744	2.425	331
8f	-4.620	-1.030	-2.343	3.590	2.277	346
8g	-4.766	-1.219	-2.506	3.547	2.260	350
8h	-4.771	-1.568	-2.563	3.203	2.208	387
8i	-4.699	-1.487	-2.419	3.212	2.280	386

Compound	E_{S_0} (eV)	E_{S_1} (eV)	E_{T_1} (eV)	$\Delta E_{S_0 \rightarrow S_1}$ (H \rightarrow L)	$\Delta E_{T_1 \rightarrow S_0}$ (H \rightarrow L)	λ_{max} (nm)
12a	-5.976	-2.851	-4.063	3.125	1.913	397
12b	-5.837	-2.700	-3.953	3.137	1.884	396
12c	-5.822	-2.710	-3.942	3.112	1.880	399
12d	-5.718	-2.699	-3.962	3.019	1.756	411
13a	-5.720	-2.610	-3.892	3.110	1.828	399
13b	-5.719	-2.608	-3.892	3.111	1.827	399
13c	-5.725	-2.610	-3.880	3.115	1.845	398
13d	-5.814	-2.584	-3.876	3.230	1.938	384
13e	-5.724	-2.609	-3.893	3.115	1.831	398
13f	-5.814	-2.584	-3.876	3.230	1.938	384
13g	-5.761	-2.665	-3.940	3.096	1.821	401

Figure 6. Energy of the ground state, singlet excited state, and triplet excited state of berberine and its analogs from TD-DFT calculations. Electronic transitions are reported in eV, and the wavelength of maximal absorbance is reported in nanometers. Calculations were performed using the B3LYP/def2-TZVP level of theory.

1 can be effectively calculated from Fermi's golden rule based
2 on the spin-orbit coupling matrix elements, but this is not
3 easily implemented in TD-DFT calculations in ORCA (27).

4 MD simulations were used to probe the potential stabilizing
5 effects and stabilization timescale of the binary complexes
6 between G4DNA and berberine, dihydroberberine, and
7 8-methylberberine. Specifically, stabilization over time of the
8 G4DNA by berberine and the two analogs were calculated
9 in GROMACS (Figure 8a), and the average RMSD was
10 calculated over the same time interval (Figure 8b). G4DNA
11 with berberine had a lower maximum RMSD value than
12 G4DNA with dihydroberberine and 8-methylberberine. RMSD
13 is a measure of the change in atom position from an initial
14 point to a final point, and a lower RMSD value indicates that
15 an atom or system has moved less from its initial position.
16 A lower RMSD value over time indicates that the complex
17 has been relatively stable. Unpaired t-testing revealed that
18 the differences in RMSD between 8-methylberberine (2-
19 tail unpaired *t*-test, *p*-value 0.10), dihydroberberine (2-
20 tail unpaired *t*-test, *p*-value 0.087), and G4DNA were insignificant
21 at a 95% confidence interval. However, the difference
22 between the RMSD of G4DNA and berberine was statistically
23 significant (2-tail unpaired *t*-test, *p*-value 0.00003), suggesting
24 a destabilization of G4DNA by berberine.

DISCUSSION

25
26
27 Molecular docking, TD-DFT excited state calculations,
28 and molecular dynamics simulations were performed to
29 investigate the SAR in berberine analogs on the impact of
30 aliphatic and aromatic side chains at C-8, 12, and 13 in the free
31 energy of binding towards dsDNA and G4DNA. While the ΔG
32 values of G4DNA appear positive, the results were predictive
33 because the most thermodynamically favorable binding pose
34 was accurately predicted. Analogs with the best free energy of
35 binding to dsDNA were the 12-substituted analogs, which had
36 ΔG values of -5.8 to -6.8 kcal/mol compared to a ΔG of -5.6
37 kcal/mol for berberine. The aromatic 12-substituted analogs
38 performed the best overall with ΔG values comparable to
39 berberine, likely due to the increased number of pi-stacking
40 interactions.

41
42 Contrary to our initial hypothesis, berberine analogs
43 with aromatic substitution did not always have the highest
44 free energy of binding to either dsDNA or G4DNA targets.
45 Rather, the effect of aryl versus aliphatic substitution on DNA
46 binding appears to be dependent on not only the nature of the
47 substituted group, but also the substituted carbon position (C-
48 8, C-12, C-13) and the nucleic acid target, as different trends
49 were observed in ΔG to dsDNA and G4DNA. Moreover, it
50 appears that a loss of the persistent cation in the isoquinolinium
51 core of berberine, as in the case of dihydroberberine and
52 any 8-alkyl or 8-aryl analog, is not necessarily detrimental
53 to DNA binding. Additionally, against initial expectations, MD
54 simulations do not seem to indicate net stabilization of the
55 G-quadruplex-ligand binary complex.

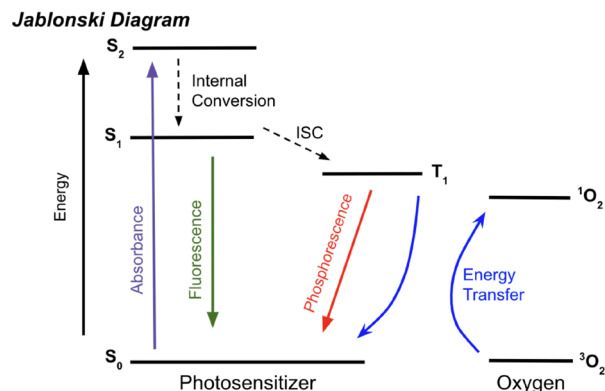
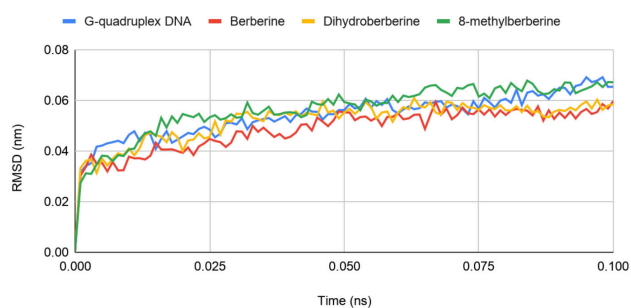


Figure 7. Jablonski Diagram depicting electronic transitions that occur as photosensitizers excite oxygen into singlet oxygen. Initial absorbance of a photon excites the ground state photosensitizer (S_0) to the second excited state (S_2), which undergoes rapid internal conversion to the first excited state (S_1). This undergoes intersystem crossing (ISC) to the first excited triplet state (T_1), which can excite ground state triplet oxygen (3O_2) to an excited singlet state (1O_2).

A



B

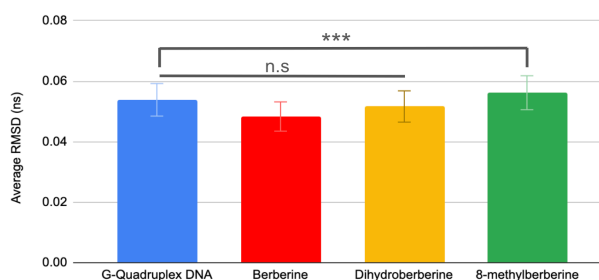


Figure 8. Root-mean-square deviation (RMSD) of atomic position calculations from molecular mechanisms. (a) Average RMSD of G4DNA, G4DNA with berberine, G4DNA with dihydroberberine, and G4DNA with 8-methylberberine over 0.1 nanosecond in water (b) Net Destabilization of G4DNA by berberine, dihydroberberine, and 8-methylberberine. 2-tail unpaired *t*-testing was completed to determine statistical significance between the average RMSD values of G4DNA compared to G4DNA with berberine, G4DNA with dihydroberberine, and G4DNA with 8-methylberberine. "n.s." indicates that the difference in average RMSD values are not statistically significant (*p*-value >0.05). "****" indicates that the difference in average RMSD values are statistically significant (*p*-value = 0.0001).

1 Visual analysis of the docked poses suggest that steric
2 effects are operative in the superior ΔG of 8-alkyl berberine
3 analogs in G4DNA (**Figure 3**, compounds **8a-8d**). The
4 crystal structure positions the compound in a manner
5 where C-8 directly faces the DNA, forcing a substituent on
6 C-8 to have more interactions with the G4DNA. However,
7 the relatively poor ΔG value of a larger substituent such as
8 the aromatic compound **8g** is potentially caused by greater
9 steric encumbrance to intercalate in G4DNA. Aliphatic
10 8-substituted analogs in the R enantiomers of G4DNA are
11 more thermodynamically favorable than S enantiomers for
12 four carbons and shorter, which can be explained by reduced
13 steric hindrance in the direction of the S enantiomers.

14 The analogs with the lowest ΔG to G4DNA and the best
15 ΔG to dsDNA were the aromatic substituents. The lower
16 affinity for aromatic chains in G4DNA can be attributed to
17 the G4DNA complex folding, exacerbating the problems of
18 steric hindrance (28). The larger variation in free energies of
19 binding with G4DNA can be attributed to the smaller binding
20 since bigger molecules, like compound **12b**, are too sterically
21 hindered to bind in more thermodynamically favorable
22 conformer poses.

23 Compound **8a** had the highest ΔG to G4DNA; however, MD
24 simulations revealed that this compound destabilized G4DNA
25 with an average RMSD value of 0.0562 ns while G4DNA
26 has an average RMSD value of 0.0538 ns. Comparatively,
27 berberine and dihydroberberine had much higher stabilization
28 capabilities with average RMSD values of 0.0483 and 0.0516
29 ns respectively, which are lower than the average G4DNA's
30 RMSD value.

31 Through TD-DFT calculations, we were able to determine
32 that the berberine analogs can undergo an electronic
33 transition that is sufficient for the production of singlet oxygen.
34 This is deemed to be beyond the current scope of the study.
35 Further studies on the lifetime of the triplet state of the
36 berberine analogs are necessary to accurately understand
37 the photosensitizing ability of berberine. One limitation with
38 the use of berberine analogs is the blueshift observed in the
39 maximum wavelength of absorbance. Many biomolecular
40 entities, such as DNA and aromatic amino acids, have
41 absorbance in the ultraviolet range, and photodynamic
42 therapy with berberine analogs could possibly result in
43 undesired side effects (29).

44 Through molecular docking, TD-DFT, and MD simulations
45 we present an exhaustive SAR of berberine and its C-8,
46 C-12, and C-13 analogs with respect to their free energy of
47 binding to both dsDNA and G4DNA. While this study primarily
48 focused on computational work and rapid *in silico* screening
49 of such compounds, it provides the basis for future work in
50 the chemical synthesis and *in vitro* evaluation of hit structures
51 and their DNA-binding efficacy.

52 MATERIALS AND METHODS

53 Molecular Mechanics Pre-Optimization

54 Avogadro, a cross-platform molecular editor, was used to

create three-dimensional computational models of berberine
and each of the studied analogs (30). Prior to density functional
theory (DFT) geometry optimization, each model was initially
optimized by molecular mechanics using the Merck Molecular
Force Field (MMFF94) to 10,000 steps. Input files for DFT
structural optimizations were created on Avogadro.

DFT Structural Optimization

Density functional theory (DFT) is a quantum mechanical
modelling method based on electron density that is used to
calculate the ground state energies of molecules and solids
(31). Here, DFT was used to calculate the most quantum
mechanically minimized molecular geometries of berberine
analogues. ORCA, an *ab initio* quantum mechanical molecular
modeling software, was used in tandem with Avogadro to
generate ORCA input files to compute the DFT optimized
structure of berberine and each analog (32-33). An implicit
conductor-like polarizable continuum (CPCM) solvation model
of water was used to simulate the conditions of an aqueous
environment. B3LYP, a hybrid functional, was chosen for
the calculation due to its low computational cost compared
to other traditional functionals, as well as its acceptance
in the scientific community for creating low parameter,
accurate results (34). The def2-SVP basis set was used as
the compounds are comprised of light main group elements.
All DFT calculations were carried out to default convergence
thresholds. Additionally, TD-DFT, with the B3LYP hybrid
functional, was used to model and calculate the excited
singlet and triplet energies of berberine and its analogs, which
is instrumental to understanding the photosensitizing ability
of berberine analogs. The def2-TZVP basis set was used
in conjunction with the RIJCOSX approximation method to
greatly accelerate the calculation with negligible decrease in
accuracy and significant reduction of computational expense.

AutoDockTools

AutoDockTools (ADT), a part of the MGLTools suite, is a
graphical user interface that allows for the preparation and
generation of coordinate files for use in AutoDock Vina (34).
To prepare the dsDNA d(CGATCG) and G4DNA moieties for
the docking procedure, ADT was used to identify the receptors
as a macromolecule, which adds Gasteiger charges to the
molecule and merges non-polar hydrogens. The search
space for the ligand was also chosen at this stage, modelled
after 3NP6 for dsDNA and 6JWD for G4DNA. Each ligand
was prepared by importing the coordinate file into ADT as
a ligand, upon which the Gasteiger charges were computed
and applied to the molecule. The identification of the torsion
tree root allowed the number of rotatable bonds to be set,
allowing for maximum conformity of the ligand to induce fit
into the specified search grid.

AutoDock Vina

AutoDock Vina is an open source molecular docking
program, allowing us to understand and model the

1 thermodynamics of ligand-protein interactions (36).
2 Configuration of Vina included the definition of the search
3 grid from ADT. Additionally, Vina was queried to generate 15
4 conformers of each ligand rather than the default 9 binding
5 modes in order to avoid omission of possible conformers.
6 The exhaustiveness value of the search was doubled from
7 the default 8 to 16 in order to generate models from more
8 computationally exhaustive methods. The validity of the
9 docking parameters used was first assessed by comparing
10 the predicted highest-affinity binding pose of berberine to
11 dsDNA d(CGTACG) with that which was previously reported
12 in its crystal structure (PDB 3NP6), and these were found to
13 be consistent.

14 UCSF Chimera/ChimeraX

15 Visual analysis to determine the accuracy of the
16 computationally determined binding mode of the berberine
17 molecule relative to that of the crystal structure shown in
18 3NP6 and 6JWD was done in UCSF Chimera and UCSF
19 ChimeraX molecular visualization programs (37-38).

20 GROMACS

21 GROMACS, or the GROningen MACHine for Chemical
22 Simulations, is a molecular dynamics (MD) package that
23 simulates interactions between proteins and ligands using
24 Newton's laws of motion (39). GROMACS was used to
25 carry out high-level molecular dynamics simulations to
26 computationally model the interactions between atoms in the
27 most thermodynamically favorable conformers of berberine,
28 dihydroberberine, and G4DNA. The AMBER99SB force field
29 and an explicit TIP3 water solvation model were used. Energy
30 minimization was conducted in order to minimize the structure
31 and remove clashes within the system. An equilibration step
32 was conducted to meet temperature and pressure constraints
33 imposed by the MD simulation. Configuration of the force field
34 using AMBER99SB parametrics and bond-charge correction
35 (BCC) charges on the ligand level was carried out on the
36 ACPYPE Web Server (40). Additional time-dependent RMSD
37 calculations to determine net stabilization of the DNA were
38 also carried out using GROMACS. MDWeb was used to
39 generate the structure of each MD simulated system in the
40 PDB format with GROMACS trajectories for visualization in
41 Chimera (41).

42 DFT, TD-DFT, molecular docking, and molecular dynamics
43 calculations were performed on a Dell PowerEdge 710 server
44 with a 24 core Intel Xeon X5660 processor at 2.80GHz and
45 32GB RAM.

46 Wavelength of Excitation

47 The minimum energy in electron-volts (eV) to excite
48 berberine and its analogs was found by determining the energy
49 difference between the lowest unoccupied molecular orbital
50 (LUMO) by the highest occupied molecular orbital (HOMO).
51 The wavelength corresponding to the HOMO-LUMO gap was
52 determined according to the following equation:

$$\lambda = \frac{hc}{E} = \frac{(6.626 \times 10^{-34} \text{ Js})(3.00 \times 10^8 \text{ m/s}) \cdot 10^9 \text{ nm/m}}{(LUMO-HOMO) \cdot 1.602 \times 10^{-19} \text{ J/eV}}$$

HOMO and LUMO orbitals were visualized on Avogadro.
Orbital energies were calculated through time-dependent
density functional theory (TD-DFT) calculations in ORCA.

ACKNOWLEDGEMENTS

We would like to acknowledge the Scripps Research
Institute and Olson Laboratory for generously providing
Autodock Vina for use in academic research. DFT and TD-
DFT calculations were performed on ORCA, an *ab-initio*,
DFT, and semi-empirical electronic structure package
developed by Frank Neese at the Max Planck Institute for
Chemical Energy Conservation, and the authors are grateful
for open source access to the software in academic research.
Molecular visualizations were produced using the UCSF
Chimera package from the Resource for Biocomputing,
Visualization, and Informatics at the University of California,
San Francisco (supported by NIH P41 RR-01081). Molecular
graphics and analyses were performed with UCSF ChimeraX,
developed by the Resource for Biocomputing, Visualization,
and Informatics at the University of California, San Francisco,
with support from National Institutes of Health R01-GM129325
and the Office of Cyber Infrastructure and Computational
Biology, National Institute of Allergy and Infectious Diseases.

The authors declare no competing conflicts of interests in
the work presented. The authors gratefully acknowledge Prof.
Robert Downing from the Department of Computer Science &
Engineering at ASDRP for his guidance with initial setup and
remote access to the server.

Received: June 11, 2020

Accepted: November 16, 2020

Published: November --, 2020

REFERENCES

1. Grycová, Lenka, *et al.* "Quaternary Protoberberine Alkaloids." *Phytochemistry*, vol. 68, no. 2, 2007, pp. 150–175., doi:10.1016/j.phytochem.2006.10.004.
2. Wang, Ye, *et al.* "The Anti-Cancer Mechanisms of Berberine: A Review." *Cancer Management and Research*, Volume 12, 2020, pp. 695–702., doi:10.2147/cmar.s242329.
3. Stermitz, F. R., *et al.* "Synergy in a Medicinal Plant: Antimicrobial Action of Berberine Potentiated by 5'-Methoxyhydrnocarpin, a Multidrug Pump Inhibitor." *Proceedings of the National Academy of Sciences*, vol. 97, no. 4, 2000, pp. 1433–1437., doi:10.1073/pnas.030540597.
4. Wang, Haoran, *et al.* "Metformin and Berberine, Two Versatile Drugs in Treatment of Common Metabolic Diseases." *Oncotarget*, vol. 9, no. 11, 2017, pp. 10135–10146., doi:10.18632/oncotarget.20807.

- 1 5. Lau, Chi-Wai, *et al.* "Cardiovascular Actions of Berberine."
2 *Cardiovascular Drug Reviews*, vol. 19, no. 3, 2006, pp.
3 234–244., doi:10.1111/j.1527-3466.2001.tb00068.x.
- 4 6. Tillhon, Micol, *et al.* "Berberine: New Perspectives for Old
5 Remedies." *Biochemical Pharmacology*, vol. 84, no. 10,
6 2012, pp. 1260–1267., doi:10.1016/j.bcp.2012.07.018.
- 7 7. Krey, A. K., and F. E. Hahn. "Berberine: Complex with
8 DNA." *Science*, vol. 166, no. 3906, 1969, pp. 755–757.,
9 doi:10.1126/science.166.3906.755.
- 10 8. Hirakawa, Kazutaka, *et al.* "The Mechanism of Guanine
11 Specific Photooxidation in the Presence of Berberine and
12 Palmatine: Activation of Photosensitized Singlet Oxygen
13 Generation through DNA-Binding Interaction." *Chemical
14 Research in Toxicology*, vol. 18, no. 10, 2005, pp. 1545–
15 1552., doi:10.1021/tx0501740.
- 16 9. Callaghan, Susan, and Mathias O. Senge. "The Good, the
17 Bad, and the Ugly – Controlling Singlet Oxygen through
18 Design of Photosensitizers and Delivery Systems for
19 Photodynamic Therapy." *Photochemical & Photobiological
20 Sciences*, vol. 17, no. 11, 2018, pp. 1490–1514., doi:10.1039/
21 c8pp00008e.
- 22 10. Ragàs, Xavier, *et al.* "Singlet Oxygen in Antimicrobial
23 Photodynamic Therapy: Photosensitizer-Dependent
24 Production and Decay in *E. Coli*." *Molecules*, vol. 18, no. 3,
25 2013, pp. 2712–2725., doi:10.3390/molecules18032712.
- 26 11. Moraca, Federica, *et al.* "Ligand Binding to Telomeric
27 G-Quadruplex DNA Investigated by Funnel-
28 Metadynamics Simulations." *Proceedings of the National
29 Academy of Sciences*, vol. 114, no. 11, 2017, doi:10.1073/
30 pnas.1612627114.
- 31 12. Spiegel, Jochen, *et al.* "The Structure and Function of
32 DNA G-Quadruplexes." *Trends in Chemistry*, vol. 2, no.
33 2, 2020, pp. 123–136., doi:10.1016/j.trechm.2019.07.002.
- 34 13. Iwasa, Kinuko, *et al.* "Antimicrobial Activity of 8-Alkyl- and
35 8-Phenyl-Substituted Berberines and Their 12-Bromo
36 Derivatives." *Journal of Natural Products*, vol. 61, no. 9,
37 1998, pp. 1150–1153., doi:10.1021/np980044+.
- 38 14. Wang, Bo, *et al.* "Syntheses and Structure–Activity
39 Relationships in Growth Inhibition Activity against Human
40 Cancer Cell Lines of 12 Substituted Berberine Derivatives."
41 *Molecules*, vol. 25, no. 8, 2020, p. 1871., doi:10.3390/
42 molecules25081871.
- 43 15. Lee, Dong-Ung, *et al.* "Effects of 13-Alkyl-Substituted
44 Berberine Alkaloids on the Expression of COX-II,
45 TNF- α , INOS, and IL-12 Production in LPS-Stimulated
46 Macrophages." *Life Sciences*, vol. 73, no. 11, 2003, pp.
47 1401–1412., doi:10.1016/s0024-3205(03)00435-1.
- 48 16. Zhang, Lei, *et al.* "Synthesis and Cytotoxicity Evaluation
49 of 13-n-Alkyl Berberine and Palmatine Analogues as
50 Anticancer Agents." *Molecules*, vol. 17, no. 10, 2012, pp.
51 11294–11302., doi:10.3390/molecules171011294.
- 52 17. Rodrigues, Catarina A. B., *et al.* "Synthesizing a Berberine
53 Derivative and Evaluating Antimicrobial Activity To
54 Reinforce with Students the Potential Significance of Small
55 Chemical Structure Changes for Biological Systems." *Journal of Chemical Education*, vol. 95, no. 3, 2018, pp. 492–495., doi:10.1021/acs.jchemed.7b00458.
18. Dhasmana, Anupam, *et al.* "High-Throughput Virtual
Screening (HTVS) of Natural Compounds and Exploration
of Their Biomolecular Mechanisms." *New Look to
Phytomedicine*, 2019, pp. 523–548., doi:10.1016/b978-0-
12-814619-4.00020-3.
19. Kumari, Madhulata, *et al.* "High Throughput Virtual
Screening to Identify Novel Natural Product Inhibitors
for MethionyltRNA-Synthetase of *Brucella Melitensis*." *Bioinformatics*, vol. 13, no. 1, 2017, pp. 8–16., doi:10.6026/97320630013008.
20. Kontoyianni, Maria. "Docking and Virtual Screening in Drug
Discovery." *Methods in Molecular Biology Proteomics for
Drug Discovery*, 2017, pp. 255–266., doi:10.1007/978-1-
4939-7201-2_18.
21. Mignon, P. "Influence of the π - π Interaction on the
Hydrogen Bonding Capacity of Stacked DNA/RNA
Bases." *Nucleic Acids Research*, vol. 33, no. 6, 2005, pp.
1779–1789., doi:10.1093/nar/gki317.
22. Ferraroni, Marta, *et al.* "X-Ray Diffraction Analyses of the
Natural Isoquinoline Alkaloids Berberine and Sanguinarine
Complexed with Double Helix DNA d(CGTACG)." *Chemical Communications*, vol. 47, no. 17, 2011, p. 4917., doi:10.1039/c1cc10971e.
23. Wang, Fei, *et al.* "Colchicine Selective Interaction with
Oncogene RET G-Quadruplex Revealed by NMR." *Chemical Communications*, vol. 56, no. 14, 2020, pp. 2099–2102., doi:10.1039/d0cc00221f.
24. Morris, Garrett M., *et al.* "AutoDock4 And AutoDockTools4:
Automated Docking with Selective Receptor Flexibility." *Journal of Computational Chemistry*, vol. 30, no. 16, 2009, pp. 2785–2791., doi:10.1002/jcc.21256.
25. Trott, Oleg, and Arthur J. Olson. "AutoDock Vina: Improving
the Speed and Accuracy of Docking with a New Scoring
Function, Efficient Optimization, and Multithreading." *Journal of Computational Chemistry*, 2009, doi:10.1002/jcc.21334.
26. Derosa, M. "Photosensitized Singlet Oxygen and Its
Applications." *Coordination Chemistry Reviews*, vol. 233-234, 2002, pp. 351–371., doi:10.1016/s0010-8545(02)00034-6.
27. Alberto, Marta Erminia, *et al.* "Rational Design of Modified
Oxobacteriochlorins as Potential Photodynamic Therapy
Photosensitizers." *International Journal of Molecular
Sciences*, vol. 20, no. 8, 2019, p. 2002., doi:10.3390/
ijms20082002.
28. Ida, Jeunice, *et al.* "G-Quadruplexes as An Alternative
Recognition Element in Disease-Related Target Sensing." *Molecules*, vol. 24, no. 6, 2019, p. 1079., doi:10.3390/molecules24061079.
29. Schmid, Franz-Xaver. "Biological Macromolecules:
UV-Visible Spectrophotometry." *Encyclopedia of Life
Sciences*, 2001, doi:10.1038/npg.els.0003142.
30. Hanwell, Marcus D, *et al.* "Avogadro: an Advanced

- 1 Semantic Chemical Editor, Visualization, and Analysis
2 Platform." *Journal of Cheminformatics*, vol. 4, no. 1, 2012,
3 doi:10.1186/1758-2946-4-17.
- 4 31. Neese, Frank. "The ORCA Program System." *WIREs*
5 *Computational Molecular Science*, vol. 2, no. 1, 2011, pp.
6 73–78., doi:10.1002/wcms.81.
- 7 32. Lee, Chengteh, *et al.* "Development of the Colle-Salvetti
8 Correlation-Energy Formula into a Functional of the
9 Electron Density." *Physical Review B*, vol. 37, no. 2, 1988,
10 pp. 785–789., doi:10.1103/physrevb.37.785.
- 11 33. Kohn, W., *et al.* "Density Functional Theory of Electronic
12 Structure." *The Journal of Physical Chemistry*, vol. 100,
13 no. 31, 1996, pp. 12974–12980., doi:10.1021/jp960669l.
- 14 34. Becke, Axel D. "Density-Functional Thermochemistry.
15 III. The Role of Exact Exchange." *The Journal of*
16 *Chemical Physics*, vol. 98, no. 7, 1993, pp. 5648–5652.,
17 doi:10.1063/1.464913.
- 18 35. Morris, Garrett M., *et al.* "AutoDock4 And AutoDockTools4:
19 Automated Docking with Selective Receptor Flexibility."
20 *Journal of Computational Chemistry*, vol. 30, no. 16, 2009,
21 pp. 2785–2791., doi:10.1002/jcc.21256.
- 22 36. Trott, Oleg, and Arthur J. Olson. "AutoDock Vina: Improving
23 the Speed and Accuracy of Docking with a New Scoring
24 Function, Efficient Optimization, and Multithreading."
25 *Journal of Computational Chemistry*, 2009, doi:10.1002/
26 jcc.21334.
- 27 37. Pettersen, Eric F., *et al.* "UCSF Chimera - A Visualization
28 System for Exploratory Research and Analysis." *Journal*
29 *of Computational Chemistry*, vol. 25, no. 13, 2004, pp.
30 1605–1612., doi:10.1002/jcc.20084.
- 31 38. Goddard, Thomas D., *et al.* "UCSF ChimeraX: Meeting
32 Modern Challenges in Visualization and Analysis." *Protein*
33 *Science*, vol. 27, no. 1, 2017, pp. 14–25., doi:10.1002/
34 pro.3235.
- 35 39. "GROMACS." *Gromacs*, www.gromacs.org/.
- 36 40. Silva, Alan W Sousa Da, and Wim F Vranken. "ACPYPE
37 - AnteChamber PYthon Parser InterfacE." *BMC Research*
38 *Notes*, vol. 5, no. 1, 2012, p. 367., doi:10.1186/1756-0500-
39 5-367.
- 40 41. Hospital, Adam, *et al.* "MDWeb and MDMoby: an
41 Integrated Web-Based Platform for Molecular Dynamics
42 Simulations." *Bioinformatics*, vol. 28, no. 9, 2012, pp.
43 1278–1279., doi:10.1093/bioinformatics/bts139.

44
45
46 **Copyright:** © 2020 Sun *et al.* All JEI articles are distributed
47 under the attribution non-commercial, no derivative license
48 (<http://creativecommons.org/licenses/by-nc-nd/3.0/>). This
49 means that anyone is free to share, copy and distribute an
50 unaltered article for non-commercial purposes provided the
51 original author and source is credited.